

University of Groningen

Erythropoietin Attenuates Pulmonary Vascular Remodeling in Experimental Pulmonary Arterial Hypertension through Interplay between Endothelial Progenitor Cells and Heme Oxygenase

van Loon, Rosa Laura E; Bartelds, Beatrijs; Wagener, Frank A D T G; Affara, Nada; Mohaupt, Saffloer; Wijnberg, Hans; Pennings, Sebastiaan W C; Takens, Janny; Berger, Rolf M F

Published in:
Frontiers in Pediatrics

DOI:
[10.3389/fped.2015.00071](https://doi.org/10.3389/fped.2015.00071)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2015

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

van Loon, R. L. E., Bartelds, B., Wagener, F. A. D. T. G., Affara, N., Mohaupt, S., Wijnberg, H., Pennings, S. W. C., Takens, J., & Berger, R. M. F. (2015). Erythropoietin Attenuates Pulmonary Vascular Remodeling in Experimental Pulmonary Arterial Hypertension through Interplay between Endothelial Progenitor Cells and Heme Oxygenase. *Frontiers in Pediatrics*, 3, [71]. <https://doi.org/10.3389/fped.2015.00071>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



Erythropoietin attenuates pulmonary vascular remodeling in experimental pulmonary arterial hypertension through interplay between endothelial progenitor cells and heme oxygenase

OPEN ACCESS

Edited by:

Maurice Beghetti,
University Hospitals of Geneva,
Switzerland

Reviewed by:

Christophe Guignabert,
University of Paris-Sud, France
Michael E. Yeager,
University of Colorado Denver, USA

*Correspondence:

Beatrijs Bartelds,
Center for Congenital Heart Diseases,
Beatrix Children's Hospital, University
Medical Center Groningen, University
of Groningen, Room W4.185, CA 41,
Hanzeplein 1, Groningen 9700 RB,
Netherlands
b.bartelds@umcg.nl

[†]Rosa Laura E. van Loon and
Beatrijs Bartelds have contributed
equally to this work.

Specialty section:

This article was submitted to Pediatric
Cardiology, a section of the journal
Frontiers in Pediatrics

Received: 04 November 2014

Accepted: 31 July 2015

Published: 28 August 2015

Citation:

van Loon RLE, Bartelds B,
Wagener FADTG, Affara N,
Mohaupt S, Wijnberg H,
Pennings SWC, Takens J and
Berger RMF (2015) Erythropoietin
attenuates pulmonary vascular
remodeling in experimental pulmonary
arterial hypertension through interplay
between endothelial progenitor cells
and heme oxygenase.
Front. Pediatr. 3:71.
doi: 10.3389/fped.2015.00071

Rosa Laura E. van Loon^{1†}, Beatrijs Bartelds^{1,2*†}, Frank A. D. T. G. Wagener³, Nada Affara¹, Saffloer Mohaupt¹, Hans Wijnberg¹, Sebastiaan W. C. Pennings³, Janny Takens^{1,2} and Rolf M. F. Berger^{1,2}

¹ Center for Congenital Heart Diseases, University Medical Center Groningen, Beatrix Children's Hospital, University of Groningen, Groningen, Netherlands, ² Department of Experimental Cardiology, University Medical Center Groningen, University of Groningen, Groningen, Netherlands, ³ Department of Orthodontics and Craniofacial Biology, Radboud University Medical Center, Nijmegen, Netherlands

Background: Pulmonary arterial hypertension (PAH) is a pulmonary vascular disease with a high mortality, characterized by typical angio-proliferative lesions. Erythropoietin (EPO) attenuates pulmonary vascular remodeling in PAH. We postulated that EPO acts through mobilization of endothelial progenitor cells (EPCs) and activation of the cytoprotective enzyme heme oxygenase-1 (HO-1).

Methods: Rats with flow-associated PAH, resembling pediatric PAH, were treated with HO-1 inducer EPO in the presence or absence of the selective HO-activity inhibitor tin-mesoporphyrin (SnMP). HO activity, circulating EPCs and pulmonary vascular lesions were assessed after 3 weeks.

Results: In PAH rats, circulating EPCs were decreased and HO activity was increased compared to control. EPO treatment restored circulating EPCs and improved pulmonary vascular remodeling, as shown by a reduced wall thickness and occlusion rate of the intra-acinar vessels. Inhibition of HO activity with SnMP aggravated PAH. Moreover, SnMP treatment abrogated EPO-induced amelioration of pulmonary vascular remodeling, while surprisingly further increasing circulating EPCs as compared with EPO alone.

Conclusion: In experimental PAH, EPO treatment restored the number of circulating EPCs to control level, improved pulmonary vascular remodeling, and showed important interplay with HO activity. Inhibition of increased HO activity in PAH rats exacerbated progression of pulmonary vascular remodeling, despite the presence of restored number of circulating EPCs. We suggest that both EPO-induced HO-1 and EPCs are promising targets to ameliorate the pulmonary vasculature in PAH.

Keywords: hemodynamics, pulmonary vascular remodeling, neointimal lesions, aorto-caval shunt, pulmonary hypertension, right ventricular hypertrophy

Introduction

Pulmonary arterial hypertension (PAH) is a life-threatening pulmonary vascular disease (1), affecting pediatric patients as well as young adults (2). Despite decades of research, pediatric patients with PAH still have a 5-year mortality rate of ~40% (2). One of the problems is that the currently available medication does not target the proliferative vascular lesions that are typical for PAH (3–5), i.e., neointimal and plexiform lesions. These lesions of the small vessel walls are formed by proliferation of endothelial and smooth muscle cells, fibrosis, thrombosis, and inflammation (4). Although the histomorphology of PAH is well described, the cellular mechanisms underlying this process are still unknown (6).

There is increasing evidence that endothelial cell dysfunction and loss of endothelial cells play a central role in the pathogenesis of pulmonary vascular remodeling (7–9). Hence, therapeutic strategies should be aimed at mobilization of endothelial cells for vascular repair (10). One of these strategies is mobilization of bone marrow-derived endothelial progenitor cells (EPCs) (10). Indeed in hypoxic pulmonary hypertension (PH), mobilization of EPC appears beneficial in reducing media hypertrophy (11). However, vascular lesions in hypoxic PH do not represent the vessel remodeling seen in PAH (3) and the effects of EPCs on vascular remodeling in neointimal proliferation are still unknown.

To study the pathogenesis and potential therapeutic strategies, we use a well-characterized animal model of flow-induced PAH, representing patients with PAH associated with congenital heart diseases (5, 6, 12). In this model, we have recently shown that erythropoietin (EPO) improves pulmonary vascular remodeling through unknown mechanisms (13). EPO is a hypoxia-induced hormone that promotes erythropoiesis. Furthermore, EPO has been demonstrated to exert therapeutic effects in various disorders affecting the vasculature, including post mechanical vascular injury (14) and myocardial and hind-limb ischemia (15, 16). These therapeutic effects may be mediated by the mobilization and homing of EPCs from the bone marrow to the injured vascular endothelium, leading to re-endothelialization and neovascularization (11, 15).

Alternatively, induction of heme oxygenase-1 (HO-1) has been implicated to play an important role in EPOs downstream mechanisms (17–19). HO-1 is the inducible isoform of heme oxygenase (HO) which catalyzes the rate-limiting step in the degradation of heme to ferrous iron, carbon monoxide, and biliverdin. Biliverdin is then rapidly converted to the powerful anti-oxidant bilirubin by biliverdin reductase (20). HO-1 is expressed in response to a variety of stresses and, in turn, its reaction products protect against oxidative and inflammatory stress and apoptosis (21, 22). In experimental models of hypoxia-induced PH, characterized by isolated media hypertrophy and not the PAH-specific obliterative pulmonary vascular remodeling, overexpression of HO-1 appears to protect against the development of PH (23, 24).

In the present study, we postulated that the beneficial effects of EPO on pulmonary vascular remodeling in PAH, are through induction of HO activity and mobilization of EPCs. To test this hypothesis, we treated rats with (flow-associated) neointimal PAH with EPO in the presence or absence the potent selective HO-activity inhibitor, tin-mesoporphyrin (SnMP), and measured

pulmonary vascular remodeling, hemodynamics, pulmonary HO activity, and the number of circulating EPCs.

Materials and Methods

Animals and Study Design

Seventy-two male Wistar rats (280–300 g) were purchased from Harlan (Zeist, The Netherlands). The experimental protocol was approved by the institutional Animal Care and Use Committee (University Medical Center Groningen). Animal care and experiments were conducted according to the US National Institutes of Health guidelines.

Rats were randomized into five groups: (1) flow-associated PAH untreated (PAH, $n = 17$), (2) PAH treated with EPO (PAH + EPO, $n = 17$), (3) PAH treated with EPO and the HO-inhibitor SnMP (PAH + EPO + SnMP, $n = 13$), (4) PAH treated with only SnMP (PAH + SnMP, $n = 12$), and (5) a control group (CON, $n = 13$). Flow-associated PAH was created by subcutaneous injection with monocrotaline (60 mg/kg, Sigma Chemical Co., St. Louis, MO, USA) at day 0, followed by an abdominal aorto-caval shunt at day 7, as described in detail previously (6). EPO (40 µg/kg, Darbepoetin-alpha, Aranesp®, Amgen Inc., Thousand Oaks, CA, USA) was administered via a single subcutaneous injection on the day of aorto-caval shunt surgery as described previously (13, 16). SnMP (30 µmol/kg, Sn(IV) Mesoporphyrin IX dichloride, Frontier Scientific Inc.) was administered via two subcutaneous injections: on the day of aorto-caval shunt surgery and 10 days later. The control group underwent subcutaneous saline injection followed by sham surgery. For analgesia, all rats received buprenorphine (0.1 mg/kg s.c. tid) for 2 days postoperatively. For measurement of hematocrit, blood (200 µL) was drawn every week starting from baseline ($t = 0$, day of monocrotaline injection). For isolation of circulating EPCs, blood (1 mL) was drawn at baseline and before sacrifice ($t = 28$).

Of the 72 rats, all but eight successfully completed the experimental protocol. In four of these eight rats, shunt surgery was not successful: two rats died due to acute surgery-related complications (PAH $n = 1$, PAH + SnMP, $n = 1$), one underwent early elective sacrifice because of postoperative paresis of hind legs (PAH) and in one rat there was no shunt present at sacrifice (PAH). Of the other four rats, one underwent early elective sacrifice because of extensive unexplained wounds to the snout (CON), one died prematurely due to unknown causes (PAH + EPO) and two rats died during orbital function for blood withdrawal (PAH + EPO, $n = 1$; PAH + SnMP, $n = 1$). All these eight rats were excluded from analysis.

Circulating Endothelial Progenitor Cells

For measuring the levels of circulating EPCs, we used a cell culturing and staining technique that has been described previously (16, 25), since fluorescence activated cell sorting (FACS) is impossible for identifying rat EPCs due to the lack of rat-specific EPC antigens.

Mononuclear cells were isolated from 1 mL of peripheral blood by density gradient centrifugation with Histopaque-1083, according to manufacturer's instructions (Sigma Chemical, St. Louis, MO, USA). Isolated mononuclear cells were seeded in triplicate on 1% gelatine-coated 96-well plates (BD BioCoat, Bedford, MA,

USA), in endothelial cell basal medium (Cambrex Bioproducts, Clonetics, NJ, USA) supplemented with 2% fetal bovine serum, EGM-2 SingleQuots (Clonetics), Penicillin (100 U/mL), and Streptomycin (100 µg/mL). After 2 days of culture, the adherent cells were carefully washed with medium and co-stained with DiI AcLDL (1,1'-diiododecyl-3,3,3',3'-tetramethylindocarbocyanine-labeled acetylated Low-Density Lipoprotein, 10 µg/mL; Molecular Probes, Invitrogen, Carlsbad, CA, USA) and FITC-conjugated BS lectin 1 (fluorescein isothiocyanate-labeled *Bandeiraea simplicifolia* lectin 1, 10 µg/mL; Sigma-Aldrich, Zwijndrecht, Netherlands). DAPI (4',6-diamidino-2-phenylindole, dilactate, Sigma-Aldrich, Zwijndrecht, Netherlands) was used for nuclear staining. Images were captured using an LSM 410 fluorescence microscope with a magnification of 100× (Carl Zeiss, Jena, Germany). Double positive cells for DiI AcLDL and lectin staining were considered an EPC and counted by co-localization analysis using Image-Pro Plus for Windows software (version 4.5) by an investigator blinded to treatment. DAPI staining served as a control to see whether the double positive cells were cells with a nucleus. For every blood sample, the average number of EPCs was calculated from six high-power field images, two from each well.

Hemodynamic Measurements

Three weeks after aorto-caval shunt surgery ($t = 28$), we anesthetized the animals and measured hemodynamics as described previously. Briefly, pulmonary and systemic arterial pressures were measured using fluid-filled catheters and right ventricular contractility was measured using a 2-French Microtip pressure transducer (Millar Instr. Inc., Houston, TX, USA). After completion of hemodynamic measurements, blood samples were drawn from the left carotid artery, distal, and proximal inferior caval vein (below and, respectively, above the level of the aorto-caval shunt) in order to determine oxygen saturation and confirm the presence of a shunt by significant oxygen step-up ($31 \pm 10\%$).

Pulmonary Vascular Remodeling

Pulmonary morphometric analysis was assessed as described previously (5, 6, 9). Briefly, the right lung was frozen in liquid nitrogen for molecular analyses. The left lung was fixated by filling the airways with 3.6% formalin at a pressure of 20 cmH₂O. Deparaffinized pulmonary 5 µm thick sections were stained with hematoxylin-eosin and Verhoef elastin-stain for morphometric analysis of vascular dimensions. In the lung sections, all transversally cut arteries with a diameter ≥ 50 µm (pre-acinar arteries) and 40 randomly chosen vessels (10 in each left lung quadrant) with a diameter < 50 µm (intra-acinar vessels) were quantitatively analyzed at 200 and 400× magnification using image analysis software (Image-Pro Plus for Windows, version 4.5). In the pre-acinar arteries, wall thickness and medial wall/lumen ratio were measured and calculated. In the intra-acinar pulmonary vessels, wall thickness was measured in addition to occlusion scores, calculated according to the following formula: (outer vessel area – luminal area)/(outer vessel area). Muscularization of 40 small pulmonary vessels was assessed as described previously (6, 12).

Heme Oxygenase Activity Measurement

To determine pulmonary HO activity at day 28, we performed an HO-activity assay by measurement of bilirubin (and biliverdin)

generation using HPLC techniques, slightly modified from previously described (26). In short, proteins were isolated from frozen lung by pulverization using a Mikro-dismembrator (Sartorius B. Braun Biotech Int., Melsungen, Germany, 2000 rpm for 40 s). Lung homogenate was resuspended (40 mg/mL) in lysis buffer (20 mM Tris/250 mM sucrose, pH = 8.5; supplemented with protease inhibitors) and subsequently lysed by three freeze-thaw cycles. Next, the lysate was centrifuged at 13,000 rpm at 4°C for 10 min. The supernatant was used for the HO-activity assay and the measured protein concentration was used for normalization. Samples for measurement on an HPLC (Spectra-Physics Analytical, Spectrasystem SCM400) equipped with a 5 µm Discovery C18 column and a Discovery C18 Supelguard Cartridge 5 µm particle size precolumn (both Sigma-Aldrich) were supplemented with vitamin C to prevent oxidation of bilirubin or biliverdin. As controls for HO activity, we used untreated fibroblasts in combination with fibroblasts treated for 24 h with 10 µM curcumin (26).

Quantitative Real-Time-PCR

Real-time PCR (RT-PCR) was performed using standard assays. Total RNA was isolated from pulmonary tissue using TRIzol reagent (Invitrogen). RT-PCR experiments were performed on a CFX384 real-time system C1000 Thermal cycler (BioRad Laboratories, Veenendaal, The Netherlands). cDNA was synthesized using Moloney murine leukemia virus reverse transcriptase and random primers (Invitrogen, Burlington, ON, Canada). RT-PCR was conducted using SYBR Green PCR Master Mix according to the manufacturer's instructions (Eurogentec, San Diego, CA, USA). Primer sequences are available on request. Expression levels were obtained from a dilution standard curve and compared with those of 36B4 RNA in order to calculate the relative expression levels.

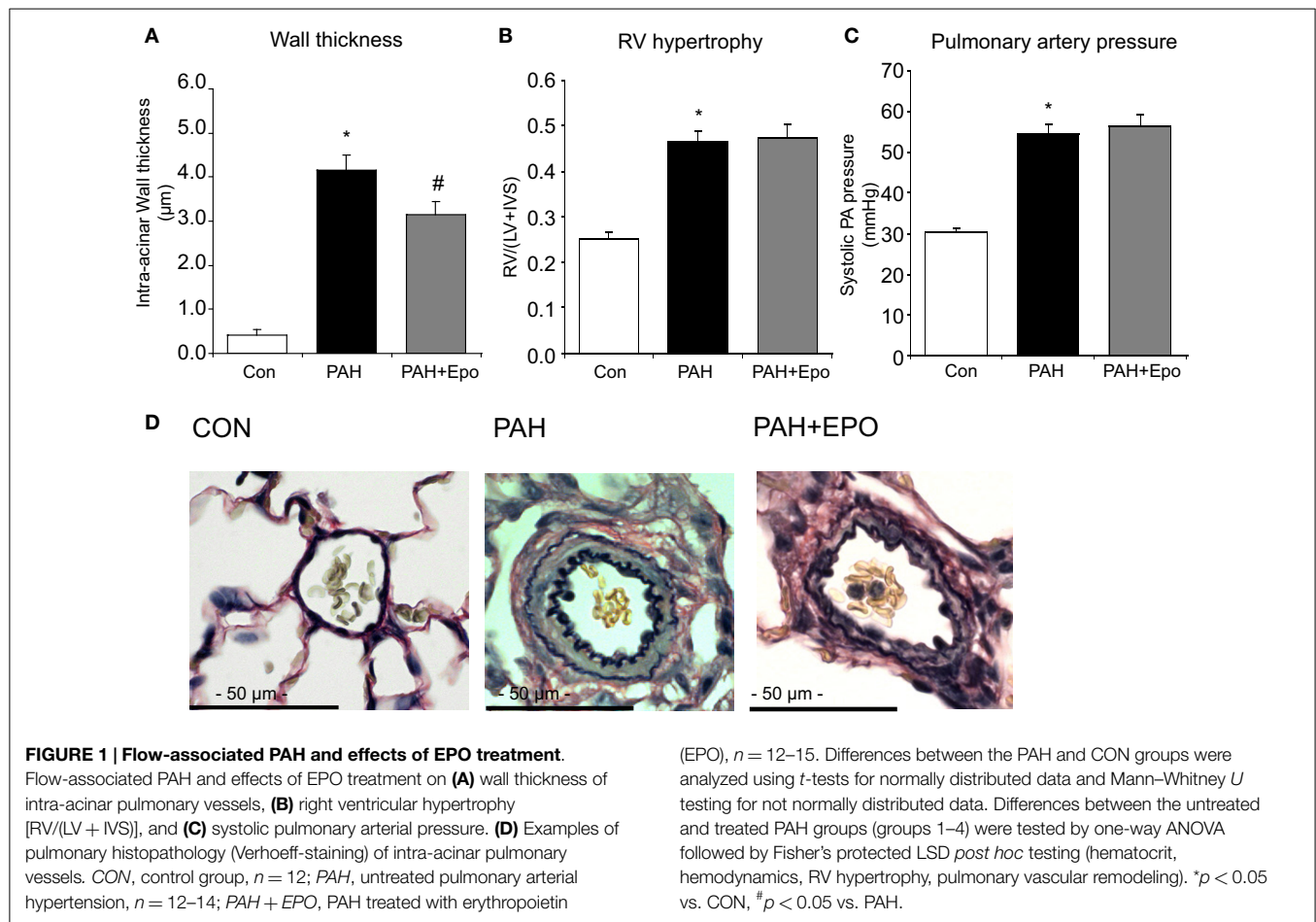
Statistical Analysis

Data are presented as mean \pm SEM or median (range). Differences between the PAH and CON groups were analyzed using *t*-tests for normally distributed data and Mann-Whitney *U* testing for not normally distributed data. Differences between the untreated and treated PAH groups (groups 1–4) were tested by one-way ANOVA followed by Fisher's protected LSD *post hoc* testing (hematocrit, hemodynamics, RV hypertrophy, and pulmonary vascular remodeling) and Kruskal-Wallis followed by Mann-Whitney *post hoc* testing with Bonferroni correction (number of EPCs) was used. Alpha was chosen to be 0.05. A two-tailed *p*-value < 0.05 was considered statistically significant. SPSS 16.0 was used to perform the analyses.

Results

Flow-Associated PAH Rat Model

Flow-associated PAH was confirmed by histology, pathology, and hemodynamics as described previously (5, 6). Briefly, PAH rats showed advanced pulmonary vascular remodeling, including significantly increased wall thickness, occlusion scores and muscularization of intra-acinar vessels as well as increased wall thickness and wall/lumen ratio of pre-acinar pulmonary vessels



as compared to control (Figures 1A,D; Table 1). PAH rats had significantly increased right ventricular hypertrophy, increased right atrial and pulmonary arterial pressures, and decreased right ventricular contractility (Figures 1B,C; Table 1).

Increased HO Activity and Decreased Number of Circulating EPCs in PAH

At day 28, rats with PAH had increased pulmonary HO activity and HO-1 mRNA levels (Figures 2A,B). The number of circulating EPCs tended to be lower in PAH, although not reaching statistical significance ($p = 0.1$, Figure 2C). There were no differences in baseline EPC number or hematocrit (data not shown).

Effects of EPO in PAH

EPO treatment improved pulmonary vascular remodeling, as previously shown by reduced wall thickness and occlusion scores of intra-acinar pulmonary vessels, compared with PAH (Figures 1A,D; Table 1). Hemodynamics remained unchanged, which is consistent with previous experiments (13). EPO treatment did not further increase pulmonary HO activity at day 28 (Figure 2A), whereas HO-1 mRNA expression levels were further increased (Figure 2B). EPO treatment increased the number of circulating EPCs to levels similar to CON (Figure 2C). Hematocrit levels increased significantly after EPO treatment (Figure 3A) demonstrating the successful

administration of EPO. Taken together, EPO improved pulmonary vascular remodeling and increased the number of circulating EPCs.

Effects of Inhibition of HO Activity in PAH

SnMP successfully inhibited HO activity in both PAH + SnMP and PAH + EPO + SnMP to levels as measured in CON (Figure 3B), whereas SnMP did not decrease HO-1 mRNA levels (Figure 3C), which confirms its function in inhibiting HO activity and not HO-1 transcription. SnMP treatment in the absence of EPO treatment aggravated pulmonary vascular remodeling to levels worse than seen in untreated PAH: significantly increased wall thickness and occlusion scores of intra-acinar pulmonary vessels and increased wall-lumen ratio of pre-acinar pulmonary arteries (Figure 3B; Table 1). Interestingly, SnMP treatment alone tended to increase circulating EPCs (PAH + SnMP vs. PAH $p = 0.09$) to a level comparable to that after EPO treatment (PAH + EPO; Figure 3B). Taken together, inhibition of HO activity worsened pulmonary vascular remodeling, despite increasing the number of circulating EPCs.

Effects of EPO and Inhibition of HO Activity in PAH

EPO-mediated amelioration of pulmonary vascular remodeling was abrogated following co-treatment with SnMP, while further

TABLE 1 | Animal characteristics.

	CON	PAH	PAH + EPO	PAH + EPO + SnMP	PAH + SnMP
Hemodynamics					
Mean pulmonary artery pressure	20 ± 1	32 ± 1*	35 ± 1	37 ± 3	34 ± 2
Systolic pulmonary arterial pressure (mmHg)	30 ± 1	54 ± 3*	56 ± 3	56 ± 4	56 ± 6
Mean systemic arterial pressure (mmHg)	89 ± 2	63 ± 4*	71 ± 6	71 ± 5	67 ± 4
dP/dT indexed max	106 ± 8	71 ± 2*	76 ± 6	74 ± 7	67 ± 6
–dP/dT indexed max	91 ± 7	55 ± 16*	80 ± 6	56 ± 18	62 ± 7
Heart rate (beats per minute)	343 ± 20	310 ± 14	321 ± 15	312 ± 18	328 ± 13
Pathology					
Body weight at sacrifice (g)	390 ± 8	350 ± 8*	346 ± 5	343 ± 5	343 ± 5
RV hypertrophy, RV/(LV + IVS)	0.25 ± 0.01	0.47 ± 0.03*	0.47 ± 0.03	0.47 ± 0.03	0.42 ± 0.04
RV weight (mg)	0.21 ± 0.02	0.44 ± 0.02	0.44 ± 0.03	0.40 ± 0.03	0.38 ± 0.03
Pulmonary vascular morphometry					
Intra-acinar pulmonary vessels <50µm					
Totally muscularized	2.6 ± 1.2	20.3 ± 3.5*	18.9 ± 2.6	24.0 ± 5.6	28.8 ± 6.1
Partly muscularized	2.7 ± 0.9	9.4 ± 2.5*	9.6 ± 2.0	17.6 ± 2.6	13.1 ± 3.0
Pre-acinar pulmonary vessels >50µm					
Wall thickness (µm)	7.3 ± 1.3	15.6 ± 1.5*	16.1 ± 1.8	15.4 ± 1.9	16.6 ± 3.1
Wall/lumen ratio	0.08 ± 0.01	0.15 ± 0.01*	0.14 ± 0.01	0.15 ± 0.02	0.24 ± 0.05 ^{§, #}

Data are presented as mean ± SEM. CON, control group, $n = 12$; PAH, untreated pulmonary arterial hypertension, $n = 14$; PAH + EPO, PAH treated with erythropoietin (EPO), $n = 15$; PAH + EPO + SnMP, PAH treated with EPO and tin-mesoporphyrin (SnMP), $n = 13$; PAH + SnMP, PAH treated with SnMP, $n = 10$; RV, right ventricle; (–)dP/dT indexed max, RV contractility, maximal rate of increase or decrease in RV pressure [calculated as the variation in pressure (P) with time (T) corrected for RV systolic pressure]; LV, left ventricle; IVS, interventricular septum. Differences between the PAH and CON groups were analyzed using t-tests for normally distributed data and Mann–Whitney U testing for not normally distributed data. Differences between the untreated and treated PAH groups (groups 1–4) were tested by one-way ANOVA followed by Fisher's protected LSD post hoc testing (hematocrit, hemodynamics, RV hypertrophy, pulmonary vascular remodeling).

* $p < 0.05$ vs. CON.

$p < 0.05$ vs. PAH.

§ $p < 0.05$ vs. PAH + EPO + SnMP.

increasing the number of circulating EPCs compared to EPO treatment alone (**Figure 3B**). EPO-induced HO-1 mRNA levels remained high in the presence of the HO-activity inhibitor SnMP (**Figure 3C**). Taken together, inhibition of HO activity strongly attenuated the protective effects of EPO on pulmonary vascular remodeling despite further increasing the number of circulating EPCs.

Effects of EPO and Inhibition of HO Activity on EPC Homing Factors and Markers of Inflammation

In order to investigate the effects of EPO on factors involved in the homing of circulating EPCs to the diseased vessel endothelium, we studied two stimuli required for EPCs to home to diseased vascular walls: vascular endothelial growth factor (VEGF) and its two receptor subtypes (VEGF-R1, VEGF-R2) and stromal derived growth factor-1 (SDF-1) (16, 27). Since inflammatory processes are involved in the pathophysiology of PAH and since HO activity has potent anti-inflammatory properties (28), we also measured monocyte chemo attractant protein-1 (MCP-1) levels, a major inflammatory marker, in order to study the effects of modulation of HO activity using EPO and SnMP on inflammatory processes in PAH.

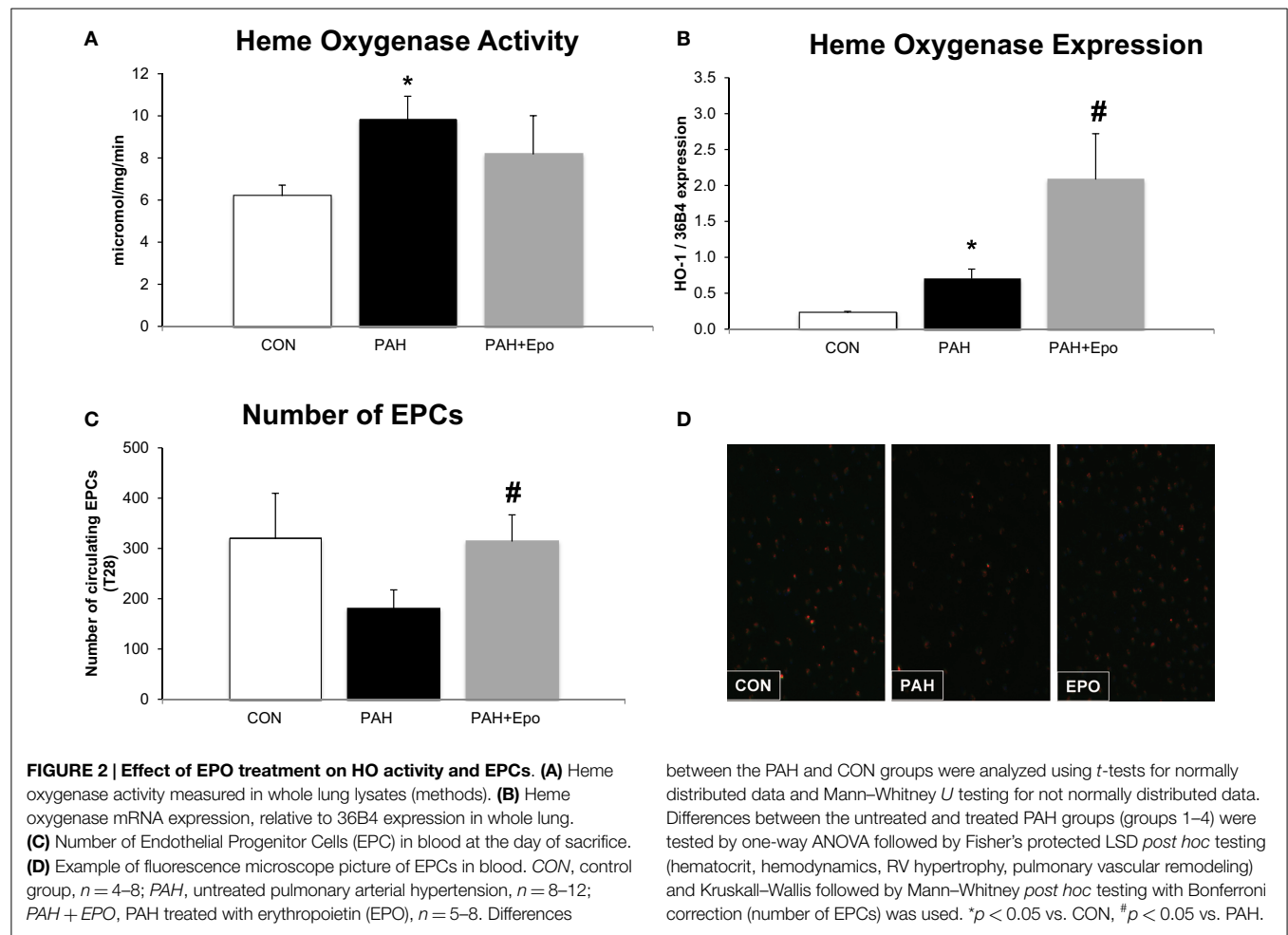
Pulmonary VEGF-R2 mRNA expression levels were decreased and SDF-1 levels were increased in PAH. After EPO treatment, these mRNA levels were unchanged (**Figure 3C**). Interestingly, EPO treatment together with SnMP tended to increase mRNA expression levels of VEGF-R2 and SDF-1, whereas SnMP alone did not alter these mRNA levels. Regarding inflammation, MCP-1

mRNA expression levels were significantly increased in PAH rats, compared to CON (**Figure 3C**). EPO treatment with or without SnMP did not influence these levels. Taken together, EPO and SnMP did not influence EPC homing factors or markers of inflammation.

Discussion

EPO treatment has been demonstrated to have beneficial effects on pulmonary vascular remodeling in PH. This study describes the interplay between EPO treatment, circulating EPCs and HO activity in a rat model of experimental flow-associated PAH. This model is characterized by a neointimal-type of pulmonary vascular remodeling, a decreased number of circulating EPCs, while both gene expression and activity of HO-1 are increased; Treatment with EPO restored the number of circulating EPCs and was associated with improved vascular remodeling. Twenty-one days after EPO administration, no change in HO activity could be demonstrated, although HO-1 gene expression was further increased. In contrast, reducing HO-1 activity to control level using tin-mesoporphyrin seriously aggravated pulmonary vascular remodeling, despite an associated increase in circulating EPCs. The combination of EPO treatment and simultaneously reduced of HO activity (to control level) resulted in a cumulative increase in circulating EPCs, however without any improvement of pulmonary vascular remodeling.

What do we learn from these observations regarding the mechanisms by which EPO treatment exerts its ameliorating effects on pulmonary vascular remodeling in PAH?



Endothelial Progenitor Cells

Consistent with previous experimental data and with recent reports in human PAH, both idiopathic and associated with congenital heart disease, we observed decreased number of circulating EPCs in untreated PAH (13, 29, 30). This shortage of circulating EPCs in PAH may contribute to the progression of pulmonary endothelial dysfunction and pulmonary vascular remodeling, since EPCs are believed to be able to incorporate into damaged endothelium and potentially repair this endothelium. Bone marrow-derived EPCs have been observed in diseased pulmonary vessel walls of experimental models of untreated hypoxia and monocrotaline-induced PH (31, 32) and the incorporation of EPCs into this diseased pulmonary endothelium has been suggested to improve pulmonary vascular remodeling (11). Until now, such improvements have been observed only after administration of genetically modified and functionally enhanced EPCs in models of media hypertrophy type of pulmonary vascular remodeling (33, 34).

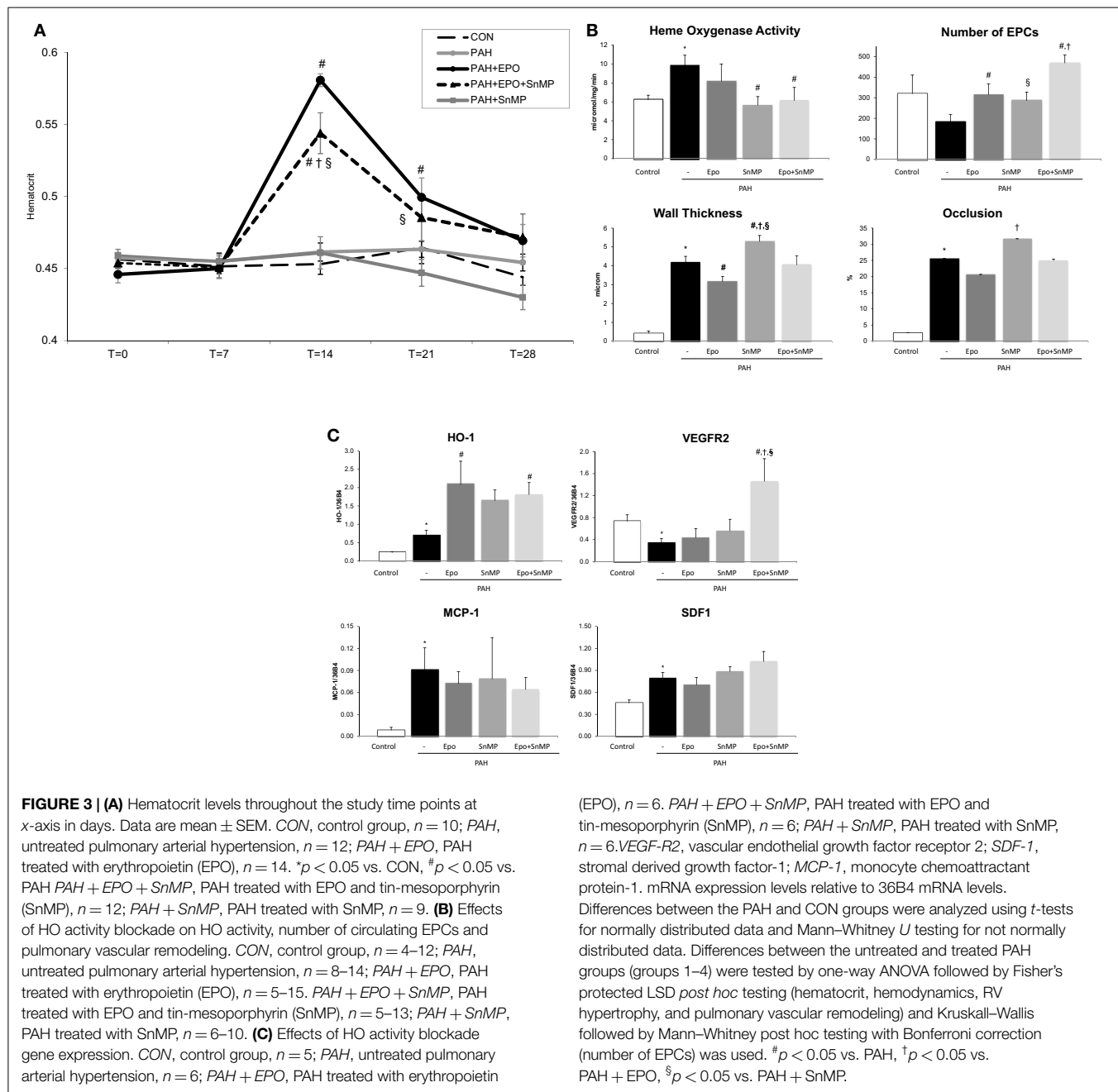
The role of bone marrow-derived progenitor cells and their tissue-restoring potential in PAH remain largely unknown and whether the beneficial effect of EPO treatment in PAH is through this mechanism remains to be demonstrated. In the current study, reducing HO activity by SnMP in experimental PAH,

induced an increase in circulating EPCs, but was associated with a further deterioration of pulmonary vascular remodeling. This observation implies that HO activity is crucially important in PAH, either for circulating EPCs to exert their healing effect on the pulmonary vasculature, or because of an EPC-independent vascular protective HO-effect, for which the increase in EPCs could not compensate. In both explanations, HO-1 appears crucial for a vascular protective effect in PAH.

Available evidence supporting the first concept that HO-1 activity is essential for EPCs to function normally and to exert the beneficial effect of EPO in PAH includes previous findings that (1) the function of EPCs and its homing to the damaged pulmonary vasculature is hampered in humans with PAH (29, 30); (2) inhibition of HO activity leads to dysfunctional EPCs (35); (3) HO activity promotes EPCs to home (36, 37); (4) beneficial effects of EPO fade out in the presence of HO-activity inhibitors in WT mice and are absent in HO-1 knock out mice (17, 19); and (5) HO-1/carbon monoxide enhances re-endothelialization after vascular injury via promoting mobilization of circulating EPCs (38).

Heme Oxygenase Activity

It is currently well established that HO-1 expression provides a general cyto- and tissue-protective effect, as a generalized



protective response to environmental stressors, including vascular shear stress. These protective effects of HO-1 are contributed to its properties to inhibit or modulate inflammatory, apoptotic and proliferative processes. HO-1 plays an essential role in such processes, via the production of the gasotransmitter carbon monoxide, the anti-oxidant bilirubin, and of iron, that co-induces the iron scavenger ferritin (21, 22). Inflammatory, apoptotic, and proliferative processes have been demonstrated to contribute to the progression of PAH (12, 28) and experimental induction of HO-1, either chemically or using transgenic mice with lung-specific HO-1-overexpression, inhibited the development of PAH in animal models (23, 24). In the present study, HO-1 mRNA and HO-1 activity levels were

augmented in untreated PAH and inhibition of HO activity worsened pulmonary vascular remodeling in the presence of normal level of circulating EPCs. This observation supports the concept that increased HO activity reflects an intrinsic protective response to increased inflammatory stress in PAH (39). Indeed, in untreated PAH rats, we found signs of increased inflammation involving highly increased levels of MCP-1, a monocyte chemoattractant and marker for inflammation, analogous to studies in humans with PAH (40). The importance of HO-1 induction in the pulmonary vascular bed is further emphasized by the recent finding that overexpression of HO-1 improves the beneficial effects of mesenchymal stromal cells in hypoxic PH (41). Putatively, HO-1 activation by specific microRNAs may initiate the

anti-inflammatory effects, as has been suggested by recent studies in ECs (42).

EPO is known to induce HO activity both in physiological and diseased states. While EPO treatment resulted in a further increase of HO-1 mRNA levels in concordance with previous findings in experimental models of kidney failure and myocardial infarction (17, 19), HO-1 activity, a more specific assessment for functional properties, was not further enhanced. The latter, however, may be explained by the timing of the experiments: EPO-mediated HO-1 induction is likely highest within the first week. After 2 weeks, most of the induced HO-1 activity may have faded. Consequently, the presented HO-activity data may be an underestimation of the initial HO-1 induction by EPO. Interestingly, other potent activators of HO-1, such as the anti-oxidant Protandim, are also successful in increasing the protective response in PAH (43). Enhancement of HO-1 may not only target the pulmonary vascular bed but also the right ventricle, which affects outcome in PAH. Protandim has been shown to target RV fibrosis (44), which is an important feature in RV dysfunction in response to increased afterload (45–48).

Recently, several studies have suggested that HO-1 and its byproduct CO also may induce cytoprotective properties by other mechanisms, then attenuating apoptosis and inflammation. HO-1 potentially impacts the regulation of autophagy, a vital cellular process, which may in part contribute to an independent cytoprotective effect of HO-1 (49).

The results of the current study show that both EPO and HO-1 have beneficial effects on pulmonary vascular remodeling in experimental PAH and both interact with circulating endothelial cells. The results further indicate that all three, EPO, EPCs, and HO-1, are potential therapeutic targets to improve pulmonary vascular remodeling in PAH. Whether intrinsic functioning and/or homing of these circulating EPCs in the pulmonary vasculature is affected by EPO or HO-1 or both cannot be concluded from the current data. Our finding that inhibition of HO activity by SnMP boosted the mobilization of EPCs is intriguing, and could be related to feedback mechanisms that sense the lack of functional EPCs homing to the vasculature or sense the need for clinical improvement. Future experiments to address this question may use chimeric PAH rats, lethally irradiated and transplanted with green fluorescent or otherwise marked bone marrow cells, to study the migration and homing of bone marrow-derived EPCs in the pulmonary endothelium and its effects on pulmonary vascular remodeling.

Since HO activity appears to be necessary for EPC-mediated repair, this suggests that EPC therapy for PAH patients could be coupled to HO-1 induction using, e.g., EPO. This is further supported by recent evidence that EPC therapy is more efficient when HIF-1 α , an inducer of HO-1-expression, is activated (50).

Several limitations should be addressed as they may have influenced the outcome of the study. At first, the design of the present study did not allow us to analyze the incorporation of EPCs within the pulmonary vascular bed, and it is therefore not possible to assess the fate of the increased number of circulating EPCs after EPO and/or HO-inhibition. The isolated mononuclear cells in our study likely contain a mixture of progenitor cells in different

developing stages; nevertheless, the majority of them are expected to be EPCs. In this study, we did not use a control group treated with SnMP alone since tissue HO-1 expression is generally low in the absence of stress and SnMP is not causing harm under these conditions. In fact, it has been given as drug to prevent baby jaundice: the uses of synthetic heme analogs (SnMP) that are competitive inhibitors of heme oxygenase, the rate-limiting enzyme in the production of bilirubin, represent a novel means of controlling severe hyperbilirubinemia in the newborn (51). This, and the observation that in the current experiment, SnMP reduced HO-1 activity to levels not lower than controls, makes clear that theoretical HO-1-depletion effects are not to be expected. Also, we used SnMP, which inhibits the activity of both HO-1 and HO-2 isoforms. Hence it is possible that some of the potent effects of SnMP could be attributed in part to inhibition of HO-2 activity. However, HO-2 is the constitutive isoform, whereas HO-1 is the inducible isoform. From the measured mRNA expression levels, one might have concluded that HO-1 was activated upon EPO treatment, but, as mentioned above, in this study we measured the actual HO activity only after 3 weeks, which is likely an underestimation of the HO activity shortly following EPO treatment. Furthermore, the effect of multiple doses of EPO was not addressed in the present study.

In conclusion, in this *in vivo* model of flow-associated PAH we showed that EPO treatment restored the number of circulating EPCs to control level, improved pulmonary vascular remodeling, and showed important interplay with HO activity. Inhibition of increased HO activity in PAH rats exacerbated progression of pulmonary vascular remodeling, despite the presence of restored number of circulating EPCs. Moreover, EPO-induced amelioration of disease was abrogated when HO activity was reduced. These findings suggest that EPO-mediated HO-1 and EPCs both provide potential therapeutic targets in this still devastating pulmonary vascular disease.

Author Contributions

RL, BB, FW, and RB contributed to the conception and design of the work, to the drafting of the work, or revising it critically for important intellectual content. RL, BB, NA, SM, HW, SP, and JT contributed to the acquisition, analysis, or interpretation of data for the work. RL, BB, FW, NA, SM, HW, SP, JT, and RB gave final approval of the version to be published and agreed to be accountable for all aspects of the work and ensured that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Acknowledgments

This study was supported by a grant from the Sebald Fund. We acknowledge the support from the Netherlands Cardiovascular Research Initiative: the Dutch Heart Foundation, Dutch Federation of University Medical Centres, the Netherlands Organisation for Health Research and development and the Royal Netherlands Academy of Sciences, CVON nr. 2012-08, PHAEDRA.

References

- Humbert M, Sitbon O, Yaici A, Montani D, O'Callaghan DS, Jais X, et al. Survival in incident and prevalent cohorts of patients with pulmonary arterial hypertension. *Eur Respir J* (2010) **36**(3):549–55. doi:10.1183/09031936.00057010
- van Loon RL, Roofthoof MT, Hillege HL, Ten Harkel AD, van Osch-Gevers M, Delhaas T, et al. Pediatric pulmonary hypertension in the Netherlands: epidemiology and characterization during the period 1991 to 2005. *Circulation* (2011) **124**(16):1755–64. doi:10.1161/CIRCULATIONAHA.110.969584
- Stenmark KR, Meyrick B, Galie N, Mooi WJ, McMurtry IF. Animal models of pulmonary arterial hypertension: the hope for etiological discovery and pharmacological cure. *Am J Physiol Lung Cell Mol Physiol* (2009) **297**(6):L1013–32. doi:10.1152/ajplung.00217.2009
- Rabinovitch M. Pathobiology of pulmonary hypertension. *Annu Rev Pathol* (2007) **2**:369–99. doi:10.1146/annurev.pathol.2.010506.092033
- Dickinson MG, Bartelds B, Molema G, Borgdorff MA, Boersma B, Takens J, et al. Egr-1 expression during neointimal development in flow-associated pulmonary hypertension. *Am J Pathol* (2011) **179**(5):2199–209. doi:10.1016/j.ajpath.2011.07.030
- van Albada ME, Bartelds B, Wijnberg H, Mohaupt S, Dickinson MG, Schoemaker RG, et al. Gene expression profile in flow-associated pulmonary arterial hypertension with neointimal lesions. *Am J Physiol Lung Cell Mol Physiol* (2010) **298**(4):L483–91. doi:10.1152/ajplung.00106.2009
- Sakao S, Tatsumi K, Voelkel NF. Endothelial cells and pulmonary arterial hypertension: apoptosis, proliferation, interaction and transdifferentiation. *Respir Res* (2009) **10**:95. doi:10.1186/1465-9921-10-95
- Ghobadi G, Bartelds B, van der Veen SJ, Dickinson MG, Brandenburg S, Berger RM, et al. Lung irradiation induces pulmonary vascular remodeling resembling pulmonary arterial hypertension. *Thorax* (2012) **67**(4):334–41. doi:10.1136/thoraxjnl-2011-200346
- Dickinson MG, Kowalski PS, Bartelds B, Borgdorff MA, van der Feen D, Sietsma H, et al. A critical role for Egr-1 during vascular remodeling in pulmonary arterial hypertension. *Cardiovasc Res* (2014) **103**(4):573–84. doi:10.1093/cvr/cvu169
- Diller GP, Thum T, Wilkins MR, Wharton J. Endothelial progenitor cells in pulmonary arterial hypertension. *Trends Cardiovasc Med* (2010) **20**(1):22–9. doi:10.1016/j.tcm.2010.03.003
- Satoh K, Kagaya Y, Nakano M, Ito Y, Ohta J, Tada H, et al. Important role of endogenous erythropoietin system in recruitment of endothelial progenitor cells in hypoxia-induced pulmonary hypertension in mice. *Circulation* (2006) **113**(11):1442–50. doi:10.1161/CIRCULATIONAHA.105.583732
- Bartelds B, van Loon RL, Mohaupt S, Wijnberg H, Dickinson MG, Boersma B, et al. Mast cell inhibition improves pulmonary vascular remodeling in pulmonary hypertension. *Chest* (2012) **141**(3):651–60. doi:10.1378/chest.11-0663
- van Albada ME, du Marchie Sarvaas GJ, Koster J, Houwertjes MC, Berger RM, Schoemaker RG. Effects of erythropoietin on advanced pulmonary vascular remodeling. *Eur Respir J* (2008) **31**(1):126–34. doi:10.1183/09031936.00035607
- Kong D, Melo LG, Mangi AA, Zhang L, Lopez-Illasaca M, Perrella MA, et al. Enhanced inhibition of neointimal hyperplasia by genetically engineered endothelial progenitor cells. *Circulation* (2004) **109**(14):1769–75. doi:10.1161/01.CIR.0000121732.85572.6F
- Heeschen C, Aicher A, Lehmann R, Fichtlscherer S, Vasa M, Urbich C, et al. Erythropoietin is a potent physiologic stimulus for endothelial progenitor cell mobilization. *Blood* (2003) **102**(4):1340–6. doi:10.1182/blood-2003-01-0223
- Westenbrink BD, Lipsic E, van der Meer P, van der Harst P, Oeseburg H, du Marchie Sarvaas GJ, et al. Erythropoietin improves cardiac function through endothelial progenitor cell and vascular endothelial growth factor mediated neovascularization. *Eur Heart J* (2007) **28**(16):2018–27. doi:10.1093/eurheartj/ehm177
- Burger D, Xiang F, Hammoud L, Lu X, Feng Q. Role of heme oxygenase-1 in the cardioprotective effects of erythropoietin during myocardial ischemia and reperfusion. *Am J Physiol Heart Circ Physiol* (2009) **296**(1):H84–93. doi:10.1152/ajpheart.00372.2008
- Calò LA, Stanic L, Davis PA, Pagnin E, Munaretto G, Fusaro M, et al. Effect of epoetin on HO-1 mRNA level and plasma antioxidants in hemodialysis patients. *Int J Clin Pharmacol Ther* (2003) **41**(5):187–92. doi:10.5414/CP41187
- Katavetin P, Inagi R, Miyata T, Shao J, Sassa R, Adler S, et al. Erythropoietin induces heme oxygenase-1 expression and attenuates oxidative stress. *Biochem Biophys Res Commun* (2007) **359**(4):928–34. doi:10.1016/j.bbrc.2007.05.207
- Dekker D, Dorresteyn MJ, Pijnenburg M, Heemskerck S, Rasing-Hoogveld A, Burger DM, et al. The bilirubin-increasing drug atazanavir improves endothelial function in patients with type 2 diabetes mellitus. *Arterioscler Thromb Vasc Biol* (2011) **31**(2):458–63. doi:10.1161/ATVBAHA.110.211789
- Loboda A, Jazwa A, Grochot-Przeczek A, Rutkowski AJ, Cisowski J, Agarwal A, et al. Heme oxygenase-1 and the vascular bed: from molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal* (2008) **10**(10):1767–812. doi:10.1089/ars.2008.2043
- Wagener FA, Volk HD, Willis D, Abraham NG, Soares MP, Adema GJ, et al. Different faces of the heme-heme oxygenase system in inflammation. *Pharmacol Rev* (2003) **55**(3):551–71. doi:10.1124/pr.55.3.5
- Christou H, Morita T, Hsieh CM, Koike H, Arkonac B, Perrella MA, et al. Prevention of hypoxia-induced pulmonary hypertension by enhancement of endogenous heme oxygenase-1 in the rat. *Circ Res* (2000) **86**(12):1224–9. doi:10.1161/01.RES.86.12.1224
- Minamino T, Christou H, Hsieh CM, Liu Y, Dhawan V, Abraham NG, et al. Targeted expression of heme oxygenase-1 prevents the pulmonary inflammatory and vascular responses to hypoxia. *Proc Natl Acad Sci U S A* (2001) **98**(15):8798–803. doi:10.1073/pnas.161272598
- Pan Y, Wang S, Yang J, Chen B, Sun Z, Ye L, et al. Interruption of CD40 Pathway improves efficacy of transplanted endothelial progenitor cells in monocrotaline induced pulmonary arterial hypertension. *Cell Physiol Biochem* (2015) **36**(2):683–96. doi:10.1159/000430130
- Scharstuhl A, Mutsaers HA, Pennings SW, Szarek WA, Russel FG, Wagener FA. Curcumin-induced fibroblast apoptosis and in vitro wound contraction are regulated by antioxidants and heme oxygenase: implications for scar formation. *J Cell Mol Med* (2009) **13**(4):712–25. doi:10.1111/j.1582-4934.2008.00339.x
- Zheng H, Fu G, Dai T, Huang H. Migration of endothelial progenitor cells mediated by stromal cell-derived factor-1 α /CXCR4 via PI3K/Akt/eNOS signal transduction pathway. *J Cardiovasc Pharmacol* (2007) **50**(3):274–80. doi:10.1097/FJC.0b013e318093ec8f
- Hassoun PM, Mouthon L, Barbera JA, Eddahibi S, Flores SC, Grimminger F, et al. Inflammation, growth factors, and pulmonary vascular remodeling. *J Am Coll Cardiol* (2009) **54**(1 Suppl):S10–9. doi:10.1016/j.jacc.2009.04.006
- Diller GP, van Eijl S, Okonko DO, Howard LS, Ali O, Thum T, et al. Circulating endothelial progenitor cells in patients with Eisenmenger syndrome and idiopathic pulmonary arterial hypertension. *Circulation* (2008) **117**(23):3020–30. doi:10.1161/CIRCULATIONAHA.108.769646
- Junhui Z, Xingxiang W, Guosheng F, Yunpeng S, Furong Z, Junzhu C. Reduced number and activity of circulating endothelial progenitor cells in patients with idiopathic pulmonary arterial hypertension. *Respir Med* (2008) **102**(7):1073–9. doi:10.1016/j.rmed.2007.12.030
- Hayashida K, Fujita J, Miyake Y, Kawada H, Ando K, Ogawa S, et al. Bone marrow-derived cells contribute to pulmonary vascular remodeling in hypoxia-induced pulmonary hypertension. *Chest* (2005) **127**(5):1793–8. doi:10.1378/chest.127.5.1793
- Spees JL, Whitney MJ, Sullivan DE, Lasky JA, Laboy M, Ylostalo J, et al. Bone marrow progenitor cells contribute to repair and remodeling of the lung and heart in a rat model of progressive pulmonary hypertension. *FASEB J* (2008) **22**(4):1226–36. doi:10.1096/fj.07-8076com
- Nagaya N, Kangawa K, Kanda M, Uematsu M, Horio T, Fukuyama N, et al. Hybrid cell-gene therapy for pulmonary hypertension based on phagocytosing action of endothelial progenitor cells. *Circulation* (2003) **108**(7):889–95. doi:10.1161/01.CIR.0000079161.56080.22
- Zhao YD, Courtman DW, Deng Y, Kugathasan L, Zhang Q, Stewart DJ. Rescue of monocrotaline-induced pulmonary arterial hypertension using bone marrow-derived endothelial-like progenitor cells: efficacy of combined cell and eNOS gene therapy in established disease. *Circ Res* (2005) **96**(4):442–50. doi:10.1161/01.RES.0000157672.70560.7b
- Tongers J, Fiedler B, König D, Kempf T, Klein G, Heineke J, et al. Heme oxygenase-1 inhibition of MAP kinases, calcineurin/NFAT signaling, and hypertrophy in cardiac myocytes. *Cardiovasc Res* (2004) **63**(3):545–52. doi:10.1016/j.cardiores.2004.04.015
- Cao YA, Wagers AJ, Karsunky H, Zhao H, Reeves R, Wong RJ, et al. Heme oxygenase-1 deficiency leads to disrupted response to acute stress in stem cells and progenitors. *Blood* (2008) **112**(12):4494–502. doi:10.1182/blood-2007-12-127621
- Deshane J, Chen S, Caballero S, Grochot-Przeczek A, Was H, Li Calzi S, et al. Stromal cell-derived factor 1 promotes angiogenesis via a heme oxygenase

- 1-dependent mechanism. *J Exp Med* (2007) **204**(3):605–18. doi:10.1084/jem.20061609
38. Lin HH, Chen YH, Yet SF, Chau LY. After vascular injury, heme oxygenase-1/carbon monoxide enhances re-endothelialization via promoting mobilization of circulating endothelial progenitor cells. *J Thromb Haemost* (2009) **7**(8):1401–8. doi:10.1111/j.1538-7836.2009.03478.x
 39. Lundvig DM, Immenschuh S, Wagener FA. Heme oxygenase, inflammation, and fibrosis: the good, the bad, and the ugly? *Front Pharmacol* (2012) **3**:81. doi:10.3389/fphar.2012.00081
 40. Sanchez O, Marcos E, Perros F, Fadel E, Tu L, Humbert M, et al. Role of endothelium-derived CC chemokine ligand 2 in idiopathic pulmonary arterial hypertension. *Am J Respir Crit Care Med* (2007) **176**(10):1041–7. doi:10.1164/rccm.200610-1559OC
 41. Wu BJ, Midwinter RG, Cassano C, Beck K, Wang Y, Changsiri D, et al. Heme oxygenase-1 increases endothelial progenitor cells. *Arterioscler Thromb Vasc Biol* (2009) **29**(10):1537–42. doi:10.1161/ATVBAHA.109.184713
 42. Heinrich EM, Dimmeler S. MicroRNAs and stem cells: control of pluripotency, reprogramming, and lineage commitment. *Circ Res* (2012) **110**(7):1014–22. doi:10.1161/CIRCRESAHA.111.243394
 43. Velmurugan K, Alam J, McCord JM, Pugazhenth S. Synergistic induction of heme oxygenase-1 by the components of the antioxidant supplement Protandim. *Free Radic Biol Med* (2009) **46**(3):430–40. doi:10.1016/j.freeradbiomed.2008.10.050
 44. Bogaard HJ, Abe K, Vonk Noordegraaf A, Voelkel NF. The right ventricle under pressure: cellular and molecular mechanisms of right-heart failure in pulmonary hypertension. *Chest* (2009) **135**(3):794–804. doi:10.1378/chest.08-0492
 45. Bartelds B, Borgdorff MA, Smit-van Oosten A, Takens J, Boersma B, Nederhoff MG, et al. Differential responses of the right ventricle to abnormal loading conditions in mice: pressure vs. volume load. *Eur J Heart Fail* (2011) **13**(12):1275–82. doi:10.1093/eurjhf/hfr134
 46. Borgdorff MA, Bartelds B, Dickinson MG, Boersma B, Weij M, Zandvoort A, et al. Sildenafil enhances systolic adaptation, but does not prevent diastolic dysfunction, in the pressure-loaded right ventricle. *Eur J Heart Fail* (2012) **14**(9):1067–74. doi:10.1093/eurjhf/hfs094
 47. Borgdorff MA, Bartelds B, Dickinson MG, van Wiechen MP, Steendijk P, de Vroomen M, et al. Sildenafil treatment in established right ventricular dysfunction improves diastolic function and attenuates interstitial fibrosis independent from afterload. *Am J Physiol Heart Circ Physiol* (2014) **307**(3):H361–9. doi:10.1152/ajpheart.00843.2013
 48. Bartelds B, Borgdorff MAJ, Berger RMF. Right ventricular adaptation in congenital heart diseases. *J Cardiovasc Dev Dis* (2014) **1**(1):83–97. doi:10.3390/jcdd1010083
 49. Surolia R, Karki S, Kim H, Yu Z, Kulkarni T, Mirov SB, et al. Heme oxygenase-1 mediated autophagy protects against pulmonary endothelial cell death and development of emphysema in cadmium treated mice. *Am J Physiol Lung Cell Mol Physiol* (2015) **309**(3):L280–92. doi:10.1152/ajplung.00097.2015
 50. Cao G, Liu C, Wan Z, Liu K, Sun H, Sun X, et al. Combined hypoxia inducible factor-1 α and homogeneous endothelial progenitor cell therapy attenuates shunt flow-induced pulmonary arterial hypertension in rabbits. *J Thorac Cardiovasc Surg* (2015). doi:10.1016/j.jtcvs.2015.05.035
 51. Drummond GS, Kappas A. Chemoprevention of severe neonatal hyperbilirubinemia. *Semin Perinatol* (2004) **28**(5):365–8. doi:10.1053/j.semperi.2004.09.004

Conflict of Interest Statement: Rosa Laura E. van Loon, Beatrijs Bartelds, Frank A. D. T. G. Wagener, Nada Affara, Saffloer Mohaupt, Hans Wijnberg, Sebastiaan W. C. Pennings, and Janny Takens have no conflict of interest to declare. Rolf M. F. Berger has performed consultancy activities for Actelion Pharmaceuticals, GlaxoSmithKline, Lilly, and United Therapeutics, and has received lecture fees from Actelion Pharmaceuticals. His institution has received grants from Actelion Pharmaceuticals and Pfizer. The Guest Associate Editor Maurice Beghetti declares that, despite having collaborated with author Rolf M. F. Berger, the review process was handled objectively and no conflict of interest exists.

Copyright © 2015 van Loon, Bartelds, Wagener, Affara, Mohaupt, Wijnberg, Pennings, Takens and Berger. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.